A Biogenetically Patterned Synthesis of the Pyrrolizidine Alkaloid Trachelanthamidine

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A synthesis of (\pm)-trachelanthamidine (5) is described using enzymes and physiological conditions; this synthesis is patterned on the likely biosynthetic pathway to pyrrolizidine alkaloids from homospermidine (1).

Experiments with isotopically labelled compounds have shown that the pyrrolizidine base, retronecine (7) is derived biosynthetically from two molecules of ornithine or arginine via putrescine and a symmetrical C₄-N-C₄ intermediate. Evidence for homospermidine (1) as this intermediate in retronecine (7) biosynthesis in Senecio isatideus plants was obtained recently by ¹⁴C-labelling experiments, and by demonstrating that homospermidine is present in these plants.² The initial steps in the conversion of homospermidine into pyrrolizidine alkaloids plausibly involve oxidation of the primary amino-groups by a diamine oxidase enzyme to generate the dialdehyde (2). This dialdehyde has been suggested3 as an intermediate in the biosynthetic pathway, since Mannich-type condensation of (2) generates 1-formylpyrrolizidine, from which 1-hydroxymethylpyrrolizidine can be obtained by reduction (Scheme 1).4 The feasibility of this proposed biosynthetic pathway from homospermidine has been demonstrated by a synthesis of (\pm) trachelanthamidine (5) under physiological conditions using diamine oxidase to carry out the initial oxidation steps, and a coupled dehydrogenase enzyme system to complete the syn-

Homospermidine (1) was prepared in 60% yield by the reaction of benzylamine with 4-chlorobutanenitrile (2 equiv.), followed by catalytic hydrogenation.⁵ The triamine (1) was incubated at 27 °C with pea seedling diamine oxidase⁶ and catalase in phosphate buffer at pH 7.⁷ After six days, the basified mixture was extracted with chloroform to yield a colourless oil. The presumed intermediates [(3) and (4)] were reduced immediately with sodium borohydride in methanol at 0 °C for 2 h. Examination of the products by thin fayer chromatography and ¹H n.m.r. spectroscopy indicated that some starting material (1) was present. This was removed by formation of the derivative of (1) with isothiocyanatobenzene in ethanol.^{2,8} G.l.c. analysis⁹ of the remaining material, and comparison with authentic samples¹⁰ indicated that the mixture was predominantly trachelanthamidine (5), [formed from

Scheme 1

the thermally more stable *exo*-aldehyde (3)] together with *ca*. 5% isoretronecanol (6). (\pm)-Trachelanthamidine (5) (40% yield) was characterised as its picrate, m.p. 169—170 °C (lit.⁹ m.p. 170—172 °C), and as (\pm)-benzoyloxymethylpyrrolizidine hydrochloride, m.p. 185—186 °C (lit.⁴ m.p. 185—185.5 °C).

The intact transformation of homospermidine (1) into (\pm)-trachelanthamidine (5) was demonstrated as follows. [1,9- 14 C₂]-Homospermidine (specific activity 14.8 μ Ci mmol $^{-1}$)² was

treated as described above to yield (\pm) -1-benzoyloxymethylpyrrolizidine hydrochloride (14.3 μ Ci mmol⁻¹). This material was converted into (\pm) -psuedoheliotridane (8) (14.4 μ Ci mmol⁻¹) by treatment with thionyl chloride followed by lithium aluminium hydride reduction.⁴ Kuhn–Roth oxidation of (8) gave acetic acid (7.4 μ Ci mmol⁻¹) as its barium salt. Schmidt degradation of the barium acetate yielded inactive barium carbonate and methylamine (7.3 μ Ci mmol⁻¹), isolated as 5-methylamino-2,4-dinitrotoluene.¹¹ Thus, 51% of the original activity from [1,9-¹⁴C₂]homospermidine is located at C-9 of trachelanthamidine (5), indicating that (1) is not broken down (*e.g.* to putrescine) before formation of trachelanthamidine.

Finally, a 'one-pot' conversion of homospermidine (1) into trachelanthamidine (5) was achieved. [1,9- 14 C₂]-Homospermidine was incubated with diamine oxidase and catalase at 27 °C in phosphate buffer at pH 7.5. After six days, liver alcohol dehydrogenase, NADH, and ethanol (as hydride donor)¹² were added and the incubation was continued at 37 °C for 24 h. Unlabelled (\pm)-trachelanthamidine was added and the mixture of bases was isolated. (\pm)-1-Benzoyloxymethylpyrrolizidine hydrochloride was crystallised to constant specific activity which indicated a 21.8% conversion of homospermidine into (\pm)-trachelanthamidine.

Thus homospermidine can be converted into (±)-trachelanthamidine by the sequence of enzymic oxidations with diamine oxidase, non-enzymic cyclisation under physiological conditions, and reduction by a coupled dehydrogenase system. The facility of these transformations supports the suggestion that such reactions are involved in the biosynthesis of pyrrolizidine alkaloids.

I thank Dr. R. B. Herbert, University of Leeds, for advice on the isolation and use of pea seedling diamine oxidase.

Received, 2nd September 1982; Com. 1054

References

- 1 D. J. Robins, Fortschr. Chem. Org. Naturst., 1982, 41, 115.
- 2 H. A. Khan and D. J. Robins, J. Chem. Soc., Chem. Commun., 1981, 554.
- 3 E.g. T. A. Geissman and D. H. G. Crout, 'Organic Chemistry of Secondary Plant Metabolism,' Freeman Cooper, San Francisco, 1969, p. 448.
- 4 N. J. Leonard and S. W. Blum, J. Am. Chem. Soc., 1960, 82, 503.
- 5 R. J. Bergeron, P. S. Burton, K. A. McGovern, and S. J. Kline, *Synthesis*, 1981, 732.
- 6 J. M. Hill, Methods Enzymol., 1971, 17B, 730.
- S. H. Hedges and R. B. Herbert, J. Chem. Res., 1979, (S) 1, (M) 413.
- 8 B. T. Golding and I. K. Nassereddin, J. Chem. Res., 1981, (S) 342, (M) 3931.
- 9 R. F. Borch and B. C. Ho, J. Org. Chem., 1977, 42, 1225.
- 10 D. J. Robins and S. Sakdarat, J. Chem. Soc., Perkin Trans. 1, 1981, 909.
- 11 D. H. G. Crout, M. H. Benn, H. Imaseki, and T. A. Geissman, *Phytochemistry*, 1966, 5, 1.
- 12 A. R. Battersby, J. Staunton, and M. C. Summers, J. Chem. Soc., Perkin Trans. 1, 1976, 1052.